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## Spotlight on Germ Cells

# Genetic aspects of testicular germ cell tumors

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**Key words:** testicular germ cell tumor, genetics, testis cancer, Y chromosome, mutations

Testicular Germ Cell Tumor (TGCT) is the most common malignant tumor in young Caucasian men with an annual increase of 3–6% in the past 50 years. Data in the literature indicate that both environmental and genetic factors acting on the primordial gonocyte/gonocyte are implicated in the etio-pathogenesis of this tumour. Genetic linkage and genome-wide analyses did not reveal a major gene effect so far, implying that multiple loci must contribute to the development of TGCTs. Only one significant genetic risk factor has been reported, the so called “gr/gr” deletion of the Y chromosome which still request further confirmation by independent studies. On the other side, the analysis of somatic genetic changes through mutation and genome imbalance analyses and expression profiling has just began to unravel the complex interaction of multiple pathways involved in TGCTs. This review focuses on genetic factors (both genomic and somatic) involved in the etiology, progression and treatment sensitivity of TGCTs.

## Introduction

Testicular Germ Cell Tumor (TGCT) is the most common malignant tumor—accounting for up to 60% of all malignancies—in young Caucasian men aged between 20 and 40 years.<sup>1,2</sup> Clinically and histologically they are divided into seminomas and nonseminomas. The worldwide incidence of TGCTs is between 6–11 per 100,000, with significant variation. It has more than doubled in the past 50 years with an annual increase of 3–6%.<sup>3,4</sup> Epidemiological associations suggest that TGCTs are associated with other reproductive disorders, including hypospadias, cryptorchidism and impaired sperm production. Therefore, these conditions may share, in part, a common aetiology and this has given rise to the term “testicular dysgenesis syndrome”.<sup>5</sup> The TDS hypothesis proposes that abnormal gonadal development (dysgenesis) along the male lineage, which can have numerous primary causes, leads secondarily to disturbed Sertoli—and Leydig cell function, resulting in reproductive disorders.

Concerning TGCTs, there is accumulating evidence of an intra-uterine phase of initiation that may involve both environmental and genetic factors acting on the primordial gonocyte/gonocyte.<sup>1,6–8</sup> This is the most likely reason that markers for embryonic germ cells, like OCT3/4 (see below), are diagnostic for the precursor lesion of TGCTs. Epidemiological studies showing striking geographical and ethnic differences largely support the TDS hypothesis.<sup>9</sup> Observations on populations migrating from countries with high incidence (Denmark) and with markedly lower incidence (Finland) to Sweden provided evidence for the importance of environmental factors acting at prenatal/perinatal period. The first generation immigrants retained the incidence as in their country of origin, whereas the second generation (born in Sweden) had a similar risk to native Swedes.<sup>10</sup> The most likely environmental chemicals acting in utero able to provoke TDS seem to be endocrine disruptors (xeno-estrogens), although direct evidence is lacking so far.<sup>11</sup>

Concerning ethnic differences, it is well known that Asian and African populations have a low risk of TGCTs and in contrast to what was observed in the Scandinavian migrating populations, men with African or Asian descent maintains their low risk even inhabiting in an area of high risk. This has been observed in African-Americans living in the USA for many generations<sup>12</sup> indicating the importance of inherited susceptibility factors as well. In fact, it demonstrates the dominance of genetics over environment in these cases. This review focuses on genetic factors involved in the aetiology, progression and treatment sensitivity of TGCTs.

## Inherited Susceptibility Factors

TGCTs can occur in a sporadic or familial manner (about 2%). Family history is among the strongest risk factors for TGCTs with a relative risk to a brother of a TGCT case of 8–10 and between fathers and sons of 4–6.<sup>10,13–16</sup> These figures are much higher than those reported for most other cancer types, which rarely exceed 4.<sup>17</sup> Other evidences for strong hereditary component come from twin studies showing a greater concordance for disease in monozygotic than in dizygotic twins<sup>18,19</sup> and from segregation analysis of familial cases suggesting a recessive mode of inheritance.<sup>20</sup> The link between bilateral TGCTs and genetic factors has been further strengthened by demonstrating that brothers and fathers of bilateral cases have 4,7 fold and 3,9 fold greater risk to develop a TGCT than those of monolateral cases.<sup>21</sup> These data clearly indicate that susceptibility genes do play a role in the aetiology of TGCTs and bilateral and familial cases are the most obvious candidates for searching inherited susceptibility factors.

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## Genetic Linkage Analysis

Thanks to genetic linkage analysis a number of cancer susceptibility alleles have been identified. Concerning TGCTs a major effort was made by the International Testicular Cancer Linkage Consortium (ITCLC) in collecting a sufficiently large set of multiple-case families for genetic linkage studies.<sup>20</sup> The first set of pedigrees ( $n = 134$ ) provided strong evidence of a susceptibility locus at Xq27 in a subset of families with disease distribution compatible with X linkage ( $HLOD = 2.01$ ) and history of at least one bilateral case ( $HLOD = 4.7$ ).<sup>22</sup> The high Heterogeneity LOD score ( $HLOD$ ) was not confirmed when subsequently 163 additional pedigrees were studied. Although a relevant gene in this region cannot be excluded, if it exists it would account only for a small proportion of TGCT susceptibility, possibly related to cryptorchidism.<sup>20</sup>

The same Consortium published the results of a genome-wide analysis on 237 TGCT pedigrees indicating six “regions of interest” at different autosomal chromosomes.<sup>23</sup> However, simulation analysis suggested that neither of these loci is likely to explain a sibling relative risk of 4.<sup>20</sup> It is therefore clear that susceptibility to TGCTs cannot be due to one major gene effect and multiple loci must contribute.

## The Precursor Lesion of TGCTs

The most likely precursor of TGCT is a gonocyte or primordial germ cell that escaped normal maturation. This lesion is known as carcinoma in situ (CIS)<sup>7</sup> or the Intratubular Germ Cell Neoplasia Unclassified (ITGCNU) according to the World health Organization (WHO).<sup>24</sup> The counterpart of CIS in the dysgenetic gonad (see below) is known as gonadoblastomas. These precursors can progress to invasive components of different histologies, including seminoma as well as various types of nonseminomas (i.e., embryonal carcinoma, teratoma, yolk sac tumor and choriocarcinoma). OCT3/4 is diagnostic to identify CIS, gonadoblastoma, seminoma and embryonal carcinoma.<sup>25,26</sup> In line with this origin of TGCTs, individuals with severe abnormalities of gonadal development associated with the intersex syndrome, currently referred to as Disorders of Sex Development (DSD)<sup>27</sup> show an increased risk for this kind of cancer, in which the Y chromosome genetic material is of crucial importance.<sup>8,28</sup>

## The Y Chromosome

The commonest structural abnormalities of the Y chromosome are microdeletions of the long arm of the Y chromosome (Yq). The Yq contain three AZF (AZoospermia Factor) regions, AZFa, b and

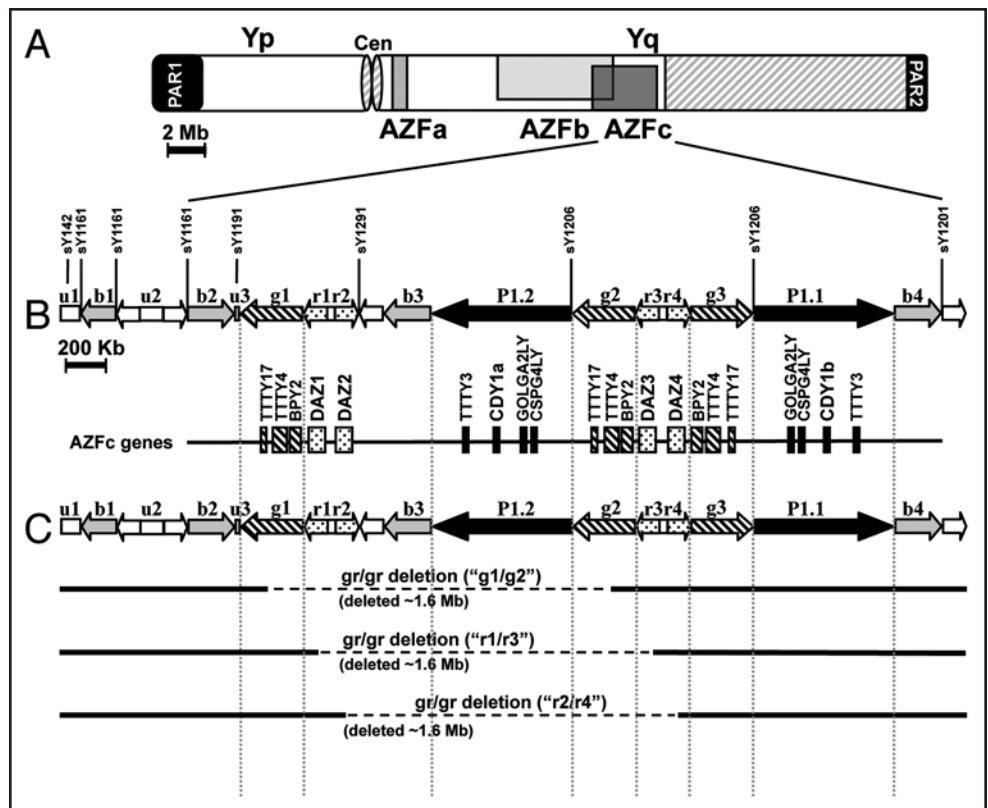


Figure 1. Schematic representation of the Y chromosome showing (A) the deletion intervals AZFa, AZFb and AZFc regions. AZFb deletions overlap with the AZFc region. Yp: short arm of the Y chromosome, cen: centromere, Yq: long arm of the Y chromosome, PAR: pseudoautosomal region. (B) the AZFc region with the indication of the location of multicopy genes and transcription units in the reference sequence published by Skaletsky et al., (2003). This region contains a number of repeated sequences with the same orientation (matching arrows) which through intrachromosomal recombination may lead to deletions. (C) gr/gr deletions which remove half of the AZFc gene content, but can vary in breakpoints. This deletion has been reported as genetic risk factor for TGCTs by Nathan et al., (2005).

c in which are mapping genes that are most likely to be involved in germ cell differentiation and spermatogenesis (Fig. 1).<sup>29,30</sup> The deletion of one or more AZF regions is strictly associated with impaired sperm production<sup>31</sup> and also distinct Y chromosome lineages (Y haplogroups) have been reported as predisposing factors for oligo/azoospermia.<sup>32,33</sup> Recently, a novel deletion designated “gr/gr”, which removes half of the gene content of the AZFc region, has been identified.<sup>34</sup> Although its clinical significance has been questioned, if only studies in which all potential methodological and selection biases were avoided are considered, “gr/gr” deletion is a significant risk factor for impaired spermatogenesis.<sup>35</sup>

While the role of Y chromosome is clearly established in male infertility, the search for Y related genetic risk factor(s) for TGCT has been largely unsuccessful. The logic behind the analysis of Y chromosome lineages in affected (cancer) versus unaffected controls is based on the assumption that an increased (or decreased) frequency of a particular Y lineage in the affected population may unmask the presence of a functional variant on the Y, in linkage with the neutral mutation defining the haplogroup. The analysis of three different ethnic groups, a total of 229 cancer patients versus 276 ethnically and geographically matched controls showed lack of association

between Y haplogroups and TGCTs.<sup>36-38</sup> Similarly, classical AZF deletions (removing all STSs located in a given AZF region) are not associated with TGCT and were not found at all in a total of 575 cancer patients.<sup>37,39-41</sup> Single STS deletions on the Y chromosome are likely to be due to PCR artefacts or to polymorphisms in the primer sequence binding site<sup>41,42</sup> which implies that they must be confirmed by sequencing the deletion breakpoints. Since confirmed single STS deletions have not been described in the literature the appealing link between Y chromosome instability (expressed as mosaic or single STS deletions) and TGCT remains to be established.

On the contrary of classical AZF deletions, “gr/gr” deletion has been reported as a significant risk factor for TGCT. A two fold increased risk of TGCT for “gr/gr” deletion carriers in 1807 affected subjects has been observed with an even higher—three fold—increase in familial cases.<sup>43</sup> In contrast to what would be expected, the authors found that probands from TGCT families exhibiting maternal lineage were at greater risk than those exhibiting paternal lineage. The association between “gr/gr” deletion and TGCT was not confirmed in a separate UK study<sup>42</sup> and an Italian study.<sup>37</sup> In general, the analysis of a very large group of subjects might be more appropriate for detecting genetic predispositions acting at low penetrance than smaller studies, however the situation is more complex for the Y chromosome than for the rest of the genome because of potential biases such as population stratification.<sup>44</sup> Ethnic and geographic matching of cases and controls is fundamental when searching for Y related factors and the multicenter study,<sup>43</sup> although very large, contained only 15% of cases and 23% of unaffected males from designed epidemiologic case-control studies. Moreover, since both gr/gr deletions and familial cases are rare the strength of the large multicenter study is still limited and would require further confirmation.

The above discussed data indicate an unlikely role for AZF deletions and genes or gene families in TGCT formation. However assuming that only a subgroup of patients will have a Y related etiology, the most likely candidates are those who present both impaired spermatogenesis and a TGCT. Only a small proportion of published studies were able to collect data on semen analysis, therefore, final conclusions are awaited.

An interesting candidate to explain the role of the Y chromosome in the development of TGCTs is the Testis Specific Protein on the Y chromosome (TSPY). This multicopy gene lies within the so called Gonadoblastoma on the Y chromosome (GBY) region, related to malignant transformation of primordial germ cell/gonocytes in patients with specific forms of DSD, especially hypovirilization and gonadal dysgenesis, and is highly expressed in CIS and GB.<sup>28,45,46</sup> Although the mechanism is still unknown, it might be related to cell cycle control.<sup>47</sup>

## Other Polymorphisms

Only a few association studies are available in the literature.<sup>6,48</sup> The two polymorphic microsatellites of the androgen receptor (*AR*) gene (CAG and GGN repeats in exon 1) have been studied in relationship with infertility,<sup>35</sup> cryptorchidism as well as TGCTs. The rationale for the analysis of *AR* polymorphisms derives from a proposed—not yet elucidated—role for androgen function in early foetal testicular development. While androgen insensitivity is an important risk factor for TGCTs, especially demonstrated in patients with DSD,<sup>28</sup> an increased androgen signalling during development

may, in theory, decrease the risk.<sup>49</sup> Given that the length of the polymorphic polyglutamin stretch (encoded by CAG repeats) influences the transactivation capacity of the receptor, variations in its length, still in the polymorphic range, may confer stronger or weaker androgenicity. Studies which focused on TGCT patients failed to find an association between CAG repeat length and cancer risk.<sup>50-52</sup> The observation by one single group about an association between long CAG tracts and tumor progression to nonseminomas and a clinically more aggressive disease awaits confirmation.<sup>51</sup>

A role in genetic susceptibility to endocrine disruption can be hypothesized for polymorphisms in those genes involved in pathways related to the action and metabolism/detoxification of endocrine disruptors or endogenous estrogens. According to the *intrauterine* origin of TGCTs a logical target for susceptibility factors is represented by maternal genetic polymorphisms. A recent case-parent study investigated a number of genes involved in the oestrogen metabolism and found an association between cytochrome *P450* gene polymorphisms (both maternal and in the index subjects) and increased risk of TGCTs.<sup>53</sup> Given the small sample size, this highly promising finding needs to be replicated by independent studies.

## The Role of Somatic Genetic Changes

**Chromosomal constitution.** Many studies investigated the chromosomal constitution of TGCTs, including its precursor lesion CIS.<sup>1,6</sup> Besides a consistent aneuploidy, specific chromosomal gains and losses are identified, but the only recurrent structural imbalance is the gain of the short arm of chromosome 12, mostly as isochromosomes. The majority of studies indicate that gain of 12p is progression related. In spite of various attempts, there is no single 12p-target gene identified. A number of genes have been suggested to be relevant, including *KRAS2*, *NANOG*, although the actual proof is lacking so far.

Single nucleotide polymorphism analysis in TGCTs demonstrated the presence of so called uniparental disomies, especially in nonseminoma.<sup>54</sup> Although of putative interest, the biological relevance remains to be shown.

Currently a number of integrated analyses of expression of genes and proteins as well as DNA copy changes are performed.<sup>1,6,55,56</sup> Overall, the data suggest a close correlation between the two, in which the expression drives the chromosomal imbalances or vice versa.

**Mutational status.** Various studies with the goal to identify pathogenetic mutations have been performed on TGCTs. Although mutations have been identified, these seem to be limited in frequency, with the possible exceptions of c-KIT and *KRAS-2*, and more recently *BRAF*. c-KIT is a kinase receptor relevant for a number of crucial processes during normal development, including survival and migration of PGCs from the epiblast to the genital ridge.<sup>57-59</sup> In normal development of germ cells, c-KIT is downregulated upon arrival of the PGCs in the genital ridge,<sup>57,58</sup> although it can still be detected at a relatively low level in human spermatogonia.<sup>60</sup> c-KIT is also present at a high level in CIS and gonadoblastoma and is overall downregulated upon invasive growth. Activating mutations, leading to a stem cell factor (SCF) independent active receptor, have been found predominantly in bilateral TGCTs but some studies found them mainly in primary unilateral seminomas.<sup>1,6</sup> The sensitivity of the mutation detection may be responsible for the conflicting data, as well as tumor progression-related loss. That indeed c-KIT has



an important role in the pathogenesis of TGCTs, is supported by the observation that this gene can be overexpressed due to a highly restricted genomic amplification only including this gene.<sup>61</sup> The most recent identification of SCF being a diagnostic marker for early malignant germ cells is of interest in this context.<sup>28</sup> The c-KIT signaling pathway has been linked to PI<sub>3</sub>K,<sup>62</sup> both in mouse PGCs as well as TGCTs. It is of interest that activating KRAS2 mutations are found in TGCTs.<sup>63,64</sup> Activation of a mutated KRAS2 results in an increased in vitro survival of seminoma cells,<sup>64,65</sup> which are normally not able to survive outside the patient, as well as an earlier age at clinical presentation of the tumor.

The proto-oncogene BRAF has been shown to be mutated in a variety of cancers, including TGCTs.<sup>66</sup> The affected pathway is the MEK-pathway, in which RAS also act. Activating mutations of KRAS and BRAF are mutually exclusive in TGCTs. A correlation between BRAF mutation and hypermethylation of the promoter of hMLH1 has been reported.<sup>67</sup> Since hMLH1 is involved in mismatch repair, absence or mutations in this gene results in micro satellite instability (MSI). Indeed, MSI instability has been reported to be related to treatment resistance (i.p. cisplatin-based) in multiple studies.<sup>68-71</sup> However, the exact link between BRAF status, MSI and treatment sensitivity of TGCTs has to be clarified.

An overall low mutation frequency is rather exceptional for solid cancers, although it seems to be the rule for TGCTs. That this is indeed not due to the pre-selection of genes under investigation, but an overall phenomenon is supported by the results of a high throughput investigation on the mutation status of the genome.<sup>72,73</sup> This might again be related to the embryonic origin of the tumors. In fact, embryonic stem cells have a unique mechanism in which one of the two DNA strands is kept protected against any form of mutations.<sup>74</sup> This protects the DNA from anomalies to be transmitted to the next generation.

**TP53 and microRNAs.** One of the intriguing observations is that also TP53 is hardly mutated in TGCTs for.<sup>1,6</sup> It is however, interesting that TP53 target genes have been found to be frequently hypermethylated in TGCTs.<sup>75</sup> The explanation for the wild type TP53 status in TGCTs was obtained, amongst others, as a result of the expression analysis of certain microRNAs.<sup>76</sup> The miRNA cluster 371–373 (mapped to chromosome 19) is specifically expressed in the seminomas and embryonal carcinomas and as expected in human embryonic stem cells.<sup>77</sup> This cluster of microRNAs was before found to be able to mimic the presence of a mutated TP53 in overruling cellular senescence in a high throughput in vitro model system.<sup>78</sup> The miRNAs interact with the 3' UTR of the mRNA encoding the tumor suppressor gene protein LATS-2, which is involved in the regulation of G<sub>1</sub>-S transition in the cell cycle. LATS-2 is indeed a downstream target of TP53, and inactivation of TP53 results in absence of LATS-2 protein, and thereby overruling cellular senescence.

## Conclusions

The analysis of familial cases did not provide evidence for linkage to any locus possibly because there are several moderate-risk genes involved in development of TGCTs. This implies that alternative approaches—not based on linkage analysis—should be taken into consideration for future investigations. One option could be the analysis of an enriched study population in specific subgroups (for example TGCT patients with impairment of spermatogenesis) versus unaffected controls. Given that spermatogenic dysfunction is more common in

TGCTs than can be explained by either local tumor or general cancer effect, it is likely that mutations in spermatogenic candidate genes will be involved only in those patients who are associated with reduced sperm count. Up to now no specific screening for mutations in spermatogenesis candidate genes has been performed in large enough group of selected cases with impaired spermatogenesis. An other even more promising alternative is a genome wide association study which could provide susceptibility locus mapping even for low penetrance disease loci. In the era of International Hapmap project (2003) and of appropriate technology platforms such studies became now a realistic way to approach multifactorial diseases. In addition, introduction of (high throughput) mutation and genome imbalance analyses and expression profiling has advanced our understanding of the pathogenic mechanisms involved in TGCTs. However, this has just began to unravel the complex interaction of multiple pathways involved, for which an integrated approach would be most informative.

## References

- Oosterhuis JW, Looijenga LH. Testicular germ-cell tumours in a broader perspective. *Nature reviews* 2005; 5:210-22.
- Ulbright TM. Germ cell neoplasms of the testis. *Am J Surg Pathol* 1993; 17:1075-91.
- Huyghe E, Matsuda T, Thonneau P. Increasing incidence of testicular cancer worldwide: a review. *The Journal of urology* 2003; 170:5-11.
- Jemal A, Tiwari RC, Murray T, Ghafoor A, Samuels A, Ward E, Feuer EJ, Thun MJ. Cancer statistics, 2004. *CA: a cancer journal for clinicians* 2004; 54:8-29.
- Skakkebaek NE, Rajpert-De Meyts E, Main KM. Testicular dysgenesis syndrome: an increasingly common developmental disorder with environmental aspects. *Hum Reprod* 2001; 16:972-8.
- Rajpert-De Meyts E. Developmental model for the pathogenesis of testicular carcinoma in situ: genetic and environmental aspects. *Hum Reprod Update* 2006; 12:303-23.
- Skakkebaek NE, Berthelsen JG, Giwercman A, Muller J. Carcinoma-in-situ of the testis: possible origin from gonocytes and precursor of all types of germ cell tumours except spermatocytoma. *Int J Androl* 1987; 10:19-28.
- Hersmus R, de Leeuw BH, Wolffenbuttel KP, Drop SL, Oosterhuis JW, Cools M, Looijenga LH. New insights into type II germ cell tumor pathogenesis based on studies of patients with various forms of disorders of sex development (DSD). *Mol Cell Endocrinol* 2008.
- McGlynn KA, Devesa SS, Graubard BI, Castle PE. Increasing incidence of testicular germ cell tumors among black men in the United States. *J Clin Oncol* 2005; 23:5757-61.
- Hemminki K, Li X. Cancer risks in Nordic immigrants and their offspring in Sweden. *Eur J Cancer* 2002; 38:2428-34.
- Sharpe RM, Skakkebaek NE. Testicular dysgenesis syndrome: mechanistic insights and potential new downstream effects. *Fertil Steril* 2008; 89:33-8.
- Gajendran VK, Nguyen M, Ellison LM. Testicular cancer patterns in African-American men. *Urology* 2005; 66:602-5.
- Heimdal K, Olsson H, Tretli S, Flodgren P, Borresen AL, Fossa SD. Familial testicular cancer in Norway and southern Sweden. *British journal of cancer* 1996; 73:964-9.
- Forman D, Oliver RT, Brett AR, Marsh SG, Moses JH, Bodmer JG, Chilvers CE, Pike MC. Familial testicular cancer: a report of the UK family register, estimation of risk and an HLA class 1 sib-pair analysis. *Br J Cancer* 1992; 65:255-62.
- Westergaard T, Olsen JH, Frisch M, Kroman N, Nielsen JW, Melbye M. Cancer risk in fathers and brothers of testicular cancer patients in Denmark. A population-based study. *International journal of cancer* 1996; 66:627-31.
- Sonneveld DJ, Sleijfer DT, Schrafford Koops H, Sijmons RH, van der Graaf WT, Sluiter WJ, Hoekstra HJ. Familial testicular cancer in a single-centre population. *Eur J Cancer* 1999; 35:1368-73.
- Dong C, Hemminki K. Modification of cancer risks in offspring by sibling and parental cancers from 2,112,616 nuclear families. *Int J Cancer* 2001; 92:144-50.
- Swerdlow AJ, De Stavola BL, Swanwick MA, Maconochie NE. Risks of breast and testicular cancers in young adult twins in England and Wales: evidence on prenatal and genetic aetiology. *Lancet* 1997; 350:1723-8.
- Lichtenstein P, Holm NV, Verkasalo PK, Iliadou A, Kaprio J, Koskenvuo M, Pukkala E, Skythe A, Hemminki K. Environmental and heritable factors in the causation of cancer—analyses of cohorts of twins from Sweden, Denmark and Finland. *The New England journal of medicine* 2000; 343:78-85.
- Rapley E. Susceptibility alleles for testicular germ cell tumour: a review. *Int J Androl* 2007; 30:242-50.
- Harland SJ, Rapley EA, Nicholson PW. Do all patients with bilateral testis cancer have a hereditary predisposition? *Int J Androl* 2007; 30:251-5.
- Rapley EA, Crockford GR, Teare D, Biggs P, Seal S, Barfoot R, Edwards S, Hamoudi R, Heimdal K, Fossa SD, Tucker K, Donald J, Collins F, Friedlander M, Hogg D, Goss P, Heidenreich A, Ormiston W, Daly PA, Forman D, Oliver TD, Leahy M, Huddart R, Cooper CS, Bodmer JG, Easton DF, Stratton MR, Bishop DT. Localization to Xq27 of a susceptibility gene for testicular germ-cell tumours. *Nature genetics* 2000; 24:197-200.

23. Crockford GP, Linger R, Hockley S, Dudakia D, Johnson L, Huddart R, Tucker K, Friedlander M, Phillips KA, Hogg D, Jewett MA, Lohynska R, Daugaard G, Richard S, Chompret A, Bonaiti-Pellie C, Heidenreich A, Albers P, Olah E, Gecki L, Bodrogi I, Ormiston WJ, Daly PA, Guilford P, Fossa SD, Heimdal K, Tjulandin SA, Liubchenko L, Stoll H, Weber W, Forman D, Oliver T, Einhorn L, McMaster M, Kramer J, Greene MH, Weber BL, Nathanson KL, Cortessis V, Easton DF, Bishop DT, Stratton MR, Rapley EA. Genome-wide linkage screen for testicular germ cell tumour susceptibility loci. *Human molecular genetics* 2006; 15:443-51.
24. Woodward PJ HA, Looijenga LHJ, et al. ed. *Testicular germ cell tumors*. Lyon: IARC Press 2004.
25. Looijenga LH, de Leeuw H, van Oorschot M, van Gurp RJ, Stoop H, Gillis AJ, de Gouveia Brazao CA, Weber RF, Kirkels WJ, van Dijk T, von Lindern M, Valk P, Lajos G, Olah E, Nesland JM, Fossa SD, Oosterhuis JW. Stem cell factor receptor (c-KIT) codon 816 mutations predict development of bilateral testicular germ-cell tumors. *Cancer Res* 2003; 63:7674-8.
26. Cheng L, Sung MT, Cossu-Rocca P, Jones TD, MacLennan GT, De Jong J, Lopez-Beltran A, Montironi R, Looijenga LH. OCT4: biological functions and clinical applications as a marker of germ cell neoplasia. *J Pathol* 2007; 211:1-9.
27. Hughes IA, Houk C, Ahmed SF, Lee PA. Consensus statement on management of intersex disorders. *Archives of disease in childhood* 2006; 91:554-63.
28. Cools M, Drop SL, Wolffenbuttel KP, Oosterhuis JW, Looijenga LH. Germ cell tumors in the intersex gonad: old paths, new directions, moving frontiers. *Endocrine reviews* 2006; 27:468-84.
29. Vogt PH, Edelmann A, Kirsch S, Henegariu O, Hirschmann P, Kieseewetter F, Kohn FM, Schill WB, Farah S, Ramos C, Hartmann M, Hartschuh W, Meschede D, Behre HM, Castel A, Nieschlag E, Weidner W, Grone HJ, Jung A, Engel W, Haidl G. Human Y chromosome azoospermia factors (AZF) mapped to different subregions in Yq11. *Hum Mol Genet* 1996; 5:933-43.
30. Skaletsky H, Kuroda-Kawaguchi T, Minx PJ, Cordum HS, Hillier L, Brown LG, Repping S, Pyntikova T, Ali J, Bieri T, Chinwalla A, Delhaunty K, Du H, Fewell G, Fulton L, Fulton R, Graves T, Hou SF, Latrielle P, Leonard S, Mardis E, Maupin R, McPherson J, Miner T, Nash W, Nguyen C, Ozersky P, Pepin K, Rock S, Rohlfing T, Scott K, Schultz B, Strong C, Tin-Wollam A, Yang SP, Waterston RH, Wilson RK, Rozen S, Page DC. The male-specific region of the human Y chromosome is a mosaic of discrete sequence classes. *Nature* 2003; 423:825-37.
31. Krausz C, Degl'Innocenti S. Y chromosome and male infertility: update, 2006. *Front Biosci* 2006; 11:3049-61.
32. Krausz C, Quintana-Murci L, Rajpert-De Meyts E, Jorgensen N, Jobling MA, Rosser ZH, Skakkebaek NE, McElreavey K. Identification of a Y chromosome haplogroup associated with reduced sperm counts. *Hum Mol Genet* 2001; 10:1873-7.
33. Yang Y, Ma M, Li L, Zhang W, Xiao C, Li S, Ma Y, Tao D, Liu Y, Lin L, Zhang S. Evidence for the association of Y-chromosome haplogroups with susceptibility to spermatogenic failure in a Chinese Han population. *J Med Genet* 2008; 45:210-5.
34. Repping S, Skaletsky H, Brown L, van Daalen SK, Korver CM, Pyntikova T, Kuroda-Kawaguchi T, de Vries JW, Oates RD, Silber S, van der Veen F, Page DC, Rozen S. Polymorphism for a 1.6-Mb deletion of the human Y chromosome persists through balance between recurrent mutation and haploid selection. *Nature genetics* 2003; 35:247-51.
35. Krausz C, Giachini C. Genetic risk factors in male infertility. *Arch Androl* 2007; 53:125-33.
36. Quintana-Murci L, Weale ME, Thomas MG, Erdei E, Bradman N, Shanks JH, Krausz C, McElreavey K. Y chromosome haplotypes and testicular cancer in the English population. *J Med Genet* 2003; 40:20.
37. Ferlin A, Speltra E, Garolla A, Selice R, Zuccarello D, Foresta C. Y chromosome haplogroups and susceptibility to testicular cancer. *Mol Hum Reprod* 2007; 13:615-9.
38. Ewis AA, Lee J, Naroda T, Kagawa S, Baba Y, Nakahori Y. Lack of association between the incidence of testicular germ cell tumors and Y-chromosome haplogroups in the Japanese population. *Int J Urol* 2006; 13:1212-7.
39. Frydelund-Larsen L, Vogt PH, Leffers H, Schadwinkel A, Daugaard G, Skakkebaek NE, Rajpert-De Meyts E. No AZF deletion in 160 patients with testicular germ cell neoplasia. *Mol Hum Reprod* 2003; 9:517-21.
40. Lutke Holzik MF, Storm K, Sijmons RH, D'Hollander M, Arts EG, Verstraaten ML, Sleijfer DT, Hoekstra HJ. Absence of constitutional Y chromosome AZF deletions in patients with testicular germ cell tumors. *Urology* 2005; 65:196-201.
41. Bor P, Hindkjaer J, Kolvraa S, Rossen P, von der Maase H, Jorgensen TM, Sorensen VT, Eiberg H, Ingwersen HJ. Screening for Y microdeletions in men with testicular cancer and undescended testis. *Journal of assisted reproduction and genetics* 2006; 23:41-5.
42. Linger R, Dudakia D, Huddart R, Easton D, Bishop DT, Stratton MR, Rapley EA. A physical analysis of the Y chromosome shows no additional deletions, other than Gr/Gr, associated with testicular germ cell tumour. *Br J Cancer* 2007; 96:357-61.
43. Nathanson KL, Kanetsky PA, Hawes R, Vaughn DJ, Letrero R, Tucker K, Friedlander M, Phillips KA, Hogg D, Jewett MA, Lohynska R, Daugaard G, Richard S, Chompret A, Bonaiti-Pellie C, Heidenreich A, Olah E, Gecki L, Bodrogi I, Ormiston WJ, Daly PA, Oosterhuis JW, Gillis AJ, Looijenga LH, Guilford P, Fossa SD, Heimdal K, Tjulandin SA, Liubchenko L, Stoll H, Weber W, Rudd M, Huddart R, Crockford GP, Forman D, Oliver DT, Einhorn L, Weber BL, Kramer J, McMaster M, Greene MH, Pike M, Cortessis V, Chen C, Schwartz SM, Bishop DT, Easton DF, Stratton MR, Rapley EA. The Y deletion gr/gr and susceptibility to testicular germ cell tumor. *Am J Hum Genet* 2005; 77:1034-43.
44. Tyler-Smith C. An evolutionary perspective on Y-chromosomal variation and male infertility. *Int J Androl* 2008.
45. Cools M, Stoop H, Kersemaekers AM, Drop SL, Wolffenbuttel KP, Bourguignon JP, Slowikowska-Hilcz J, Kula K, Faradz SM, Oosterhuis JW, Looijenga LH. Gonadoblastoma arising in undifferentiated gonadal tissue within dysgenetic gonads. *J Clin Endocrinol Metab* 2006; 91:2404-13.
46. Looijenga LH, Hersmus R, Oosterhuis JW, Cools M, Drop SL, Wolffenbuttel KP. Tumor risk in disorders of sex development (DSD). *Best practice & research* 2007; 21:480-95.
47. Oram SW, Liu XX, Lee TL, Chan WY, Lau YF. TSPY potentiates cell proliferation and tumorigenesis by promoting cell cycle progression in HeLa and NIH3T3 cells. *BMC cancer* 2006; 6:154.
48. Lutke Holzik MF, Rapley EA, Hoekstra HJ, Sleijfer DT, Nolte IM, Sijmons RH. Genetic predisposition to testicular germ-cell tumours. *Lancet Oncol* 2004; 5:363-71.
49. Rajpert-De Meyts E, Skakkebaek NE. The possible role of sex hormones in the development of testicular cancer. *Eur Urol* 1993; 23:54-9.
50. Garolla A, Ferlin A, Vinanzi C, Roverato A, Sotti G, Artibani W, Foresta C. Molecular analysis of the androgen receptor gene in testicular cancer. *Endocrine-related cancer* 2005; 12:645-55.
51. Giwercman A, Lundin KB, Eberhard J, Stahl O, Cwikiel M, Cavallin-Stahl E, Giwercman YL. Linkage between androgen receptor gene CAG trinucleotide repeat length and testicular germ cell cancer histological type and clinical stage. *Eur J Cancer* 2004; 40:2152-8.
52. Rajpert-De Meyts E, Leffers H, Daugaard G, Andersen CB, Petersen PM, Hinrichsen J, Pedersen LG, Skakkebaek NE. Analysis of the polymorphic CAG repeat length in the androgen receptor gene in patients with testicular germ cell cancer. *Int J Cancer* 2002; 102:201-4.
53. Starr JR, Chen C, Doody DR, Hsu L, Ricks S, Weiss NS, Schwartz SM. Risk of testicular germ cell cancer in relation to variation in maternal and offspring cytochrome p450 genes involved in catechol estrogen metabolism. *Cancer Epidemiol Biomarkers Prev* 2005; 14:2183-90.
54. Lu YJ, Yang J, Noel E, Skoulakis S, Chaplin T, Raghavan M, Purkis T, McIntyre A, Kudahetti SC, Naase M, Berney D, Shipley J, Oliver RT, Young BD. Association between large-scale genomic homozygosity without chromosomal loss and nonseminomatous germ cell tumor development. *Cancer Res* 2005; 65:9137-41.
55. McIntyre A, Summersgill B, Lu YJ, Missiaglia E, Kitazawa S, Oosterhuis JW, Looijenga LH, Shipley J. Genomic copy number and expression patterns in testicular germ cell tumours. *Br J Cancer* 2007; 97:1707-12.
56. Rajpert-De Meyts E. Recent advances and future directions in research on testicular germ cell cancer. *Int J Androl* 2007; 30:192-7.
57. Godin I, Deed R, Cooke J, Zsebo K, Dexter M, Wylie CC. Effects of the steel gene product on mouse primordial germ cells in culture. *Nature* 1991; 352:807-9.
58. Wylie CC. The biology of primordial germ cells. *Eur Urol* 1993; 23:62-7.
59. Runyan C, Schaible K, Molyneux K, Wang Z, Levin L, Wylie C. Steel factor controls midline cell death of primordial germ cells and is essential for their normal proliferation and migration. *Development* 2006; 133:4861-9.
60. Stoop H HF, Van de Geijn GJM, Gillis AJM, Cools MC, De Boer C, Bokemeyer C, Wolffenbuttel KP, Drop SL, De Krijger RR, Dennis N, Summersgill B, McIntyre A, Shipley J, Oosterhuis JW, Looijenga LHJ. Stem cell factor as a novel diagnostic marker for early malignant germ cells. *J Pathol* 2008; in press.
61. McIntyre A, Summersgill B, Grygalewicz B, Gillis AJ, Stoop J, van Gurp RJ, Dennis N, Fisher C, Huddart R, Cooper C, Clark J, Oosterhuis JW, Looijenga LH, Shipley J. Amplification and overexpression of the KIT gene is associated with progression in the seminoma subtype of testicular germ cell tumors of adolescents and adults. *Cancer Res* 2005; 65:8085-9.
62. De Miguel MP, Cheng L, Holland EC, Federspiel MJ, Donovan PJ. Dissection of the c-Kit signaling pathway in mouse primordial germ cells by retroviral-mediated gene transfer. *Proc Natl Acad Sci USA* 2002; 99:10458-63.
63. Goddard NC, McIntyre A, Summersgill B, Gilbert D, Kitazawa S, Shipley J. KIT and RAS signalling pathways in testicular germ cell tumours: new data and a review of the literature. *Int J Androl* 2007; 30:337-48.
64. Olie RA, Looijenga LH, Boerrigter L, Top B, Rodenhuis S, Langeveld A, Mulder MP, Oosterhuis JW. N- and KRAS mutations in primary testicular germ cell tumors: incidence and possible biological implications. *Genes Chromosomes Cancer* 1995; 12:110-6.
65. Olie RA, Looijenga LH, Dekker MC, de Jong FH, van Dissel-Emiliani FM, de Rooij DG, van der Holt B, Oosterhuis JW. Heterogeneity in the in vitro survival and proliferation of human seminoma cells. *Br J Cancer* 1995; 71:13-7.
66. Sommerer F, Hengge UR, Markwarth A, Vomschloss S, Stolzenburg JU, Wittekind C, Tannapfel A. Mutations of BRAF and RAS are rare events in germ cell tumours. *International journal of cancer* 2005; 113:329-35.
67. Imai K, Yamamoto H. Carcinogenesis and microsatellite instability: The interrelationship between genetics and epigenetics. *Carcinogenesis* 2007.
68. Mayer F, Gillis AJM, Dinjens W, Oosterhuis JW, Bokemeyer C, Looijenga LHJ. Microsatellite instability of germ cell tumors is associated with resistance to systemic treatment. *Cancer Res* 2002; 62:2758-60.
69. Devouassoux-Shisheboran M, Mauduit C, Bouvier R, Berger F, Bouras M, Droz JP, Benahmed M. Expression of hMLH1 and hMSH2 and assessment of microsatellite instability in testicular and mediastinal germ cell tumours. *Mol Hum Reprod* 2001; 7:1099-105.
70. Velasco A, Riquelme E, Schultz M, Wistuba II, Villarreal L, Koh MS, Leach FS. Microsatellite Instability and Loss of Heterozygosity Have Distinct Prognostic Value for Testicular Germ Cell Tumor Recurrence. *Cancer Biol Ther* 2004; 3.

71. Velasco A, Corvalan A, Wistuba II, Riquelme E, Chuaqui R, Majerson A, Leach FS. Mismatch repair expression in testicular cancer predicts recurrence and survival. *Int J Cancer* 2007.
72. Bignell G, Smith R, Hunter C, Stephens P, Davies H, Greenman C, Teague J, Butler A, Edkins S, Stevens C, O'Meara S, Parker A, Avis T, Barthorpe S, Brackenbury L, Buck G, Clements J, Cole J, Dicks E, Edwards K, Forbes S, Gorton M, Gray K, Halliday K, Harrison R, Hills K, Hinton J, Jones D, Kosmidou V, Laman R, Lugg R, Menzies A, Perry J, Petty R, Raine K, Shepherd R, Small A, Solomon H, Stephens Y, Tofts C, Varian J, Webb A, West S, Widaa S, Yates A, Gillis AJ, Stoop HJ, van Gurp RJ, Oosterhuis JW, Looijenga LH, Futreal PA, Wooster R, Stratton MR. Sequence analysis of the protein kinase gene family in human testicular germ-cell tumors of adolescents and adults. *Genes Chromosomes Cancer* 2006; 45:42-6.
73. Greenman C, Stephens P, Smith R, Dalgliesh GL, Hunter C, Bignell G, Davies H, Teague J, Butler A, Stevens C, Edkins S, O'Meara S, Vastrik I, Schmidt EE, Avis T, Barthorpe S, Bhamra G, Buck G, Choudhury B, Clements J, Cole J, Dicks E, Forbes S, Gray K, Halliday K, Harrison R, Hills K, Hinton J, Jenkinson A, Jones D, Menzies A, Mironenko T, Perry J, Raine K, Richardson D, Shepherd R, Small A, Tofts C, Varian J, Webb T, West S, Widaa S, Yates A, Cahill DP, Louis DN, Goldstraw P, Nicholson AG, Brasseur F, Looijenga L, Weber BL, Chiew YE, DeFazio A, Greaves MF, Green AR, Campbell P, Birney E, Easton DF, Chenevix-Trench G, Tan MH, Khoo SK, Teh BT, Yuen ST, Leung SY, Wooster R, Futreal PA, Stratton MR. Patterns of somatic mutation in human cancer genomes. *Nature* 2007; 446:153-8.
74. Hong Y, Stambrook PJ. Restoration of an absent G<sub>1</sub> arrest and protection from apoptosis in embryonic stem cells after ionizing radiation. *Proc Natl Acad Sci USA* 2004; 101:14443-8.
75. Christoph F, Kempkensteffen C, Weikert S, Krause H, Schostak M, Miller K, Schrader M. Frequent epigenetic inactivation of p53 target genes in seminomatous and nonseminomatous germ cell tumors. *Cancer Lett* 2007; 247:137-42.
76. Gillis AJ, Stoop HJ, Hersmus R, Oosterhuis JW, Sun Y, Chen C, Guenther S, Sherlock J, Veltman I, Baeten J, van der Spek PJ, de Alarcon P, Looijenga LH. High-throughput microRNAome analysis in human germ cell tumours. *J Pathol* 2007; 213:319-28.
77. Suh MR, Lee Y, Kim JY, Kim SK, Moon SH, Lee JY, Cha KY, Chung HM, Yoon HS, Moon SY, Kim VN, Kim KS. Human embryonic stem cells express a unique set of microRNAs. *Dev Biol* 2004; 270:488-98.
78. Voorhoeve PM, le Sage C, Schrier M, Gillis AJ, Stoop H, Nagel R, Liu YP, van Duijse J, Drost J, Griekspoor A, Zlotorynski E, Yabuta N, De Vita G, Nojima H, Looijenga LH, Agami R. A genetic screen implicates miRNA-372 and miRNA-373 as oncogenes in testicular germ cell tumors. *Cell* 2006; 124:1169-81.